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		Application No.		Applicant(s)			
Office Action Summary		10/049,887		CHIBA ET AL.			
		Examiner		Art Unit	·		
		Yong D. Pak		1652			
The MAILING DATE of this communication appears on the cover sheet with the correspondence address Period for Reply							
A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION. - Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication. - If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication. - Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).							
Status							
1)🛛	Responsive to communication(s) filed on 29 January 2007.						
2a)) This action is FINAL . 2b) ▼ This action is non-final.						
3)	Since this application is in condition for allowance except for formal matters, prosecution as to the merits is						
	closed in accordance with the practice under E	x parte Quayle, 1	935 C.D. 11, 45	53 O.G. 213.			
Dispositi	on of Claims						
 4) Claim(s) 88-105 is/are pending in the application. 4a) Of the above claim(s) 88-91 and 95-105 is/are withdrawn from consideration. 5) Claim(s) is/are allowed. 6) Claim(s) 88 and 92-94 is/are rejected. 7) Claim(s) is/are objected to. 8) Claim(s) are subject to restriction and/or election requirement. 							
Applicati	on Papers				•		
 9) The specification is objected to by the Examiner. 10) The drawing(s) filed on is/are: a) accepted or b) objected to by the Examiner. Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a). Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d). 11) The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152. 							
Priority u	ınder 35 U.S.C. § 119	•			•		
 12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f). a) All b) Some * c) None of: 1. Certified copies of the priority documents have been received. 2. Certified copies of the priority documents have been received in Application No 3. Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)). * See the attached detailed Office action for a list of the certified copies not received. 							
2) Notice 3) Information	t(s) se of References Cited (PTO-892) se of Draftsperson's Patent Drawing Review (PTO-948) mation Disclosure Statement(s) (PTO/SB/08) or No(s)/Mail Date	F 1 [] (5	nterview Summary Paper No(s)/Mail Da Notice of Informal P Other:	ate			

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DETAILED ACTION

Continued Examination Under 37 CFR 1.114

A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on January 29, 2007 has been entered.

Claims 88-105 are pending. Claims 89-91 and 95-105 are withdrawn. Claims 88 and 92-94 are under consideration.

Claim Objections

Claim 88 is objected to because of the following informalities:

Claim 88 recites "A <u>in vivo</u>". It appears applicants have meant to recite "An<u>in</u> vivo". Appropriate correction is required.

Claim Rejections - 35 USC § 112

The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

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Claim 88 and claims 92-94 depending therefrom are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 88 recites the term "<u>in vivo</u> method for preparing a mutant yeast". The metes and bounds of the term in the context of the claim are not clear. It is not clear to the Examiner how a method of preparing a mutant yeast can be carried out *in vivo* or how an "<u>in vivo</u> method" differs from "A method for preparing a mutant yeast..". For examination purposes, Examiner has given no weight to the term "<u>in vivo</u>".

The following is a quotation of the first paragraph of 35 U.S.C. 112:

The specification shall contain a written description of the invention, and of the manner and process of making and using it, in such full, clear, concise, and exact terms as to enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the same and shall set forth the best mode contemplated by the inventor of carrying out his invention.

Claims 88 and 92-94 remain rejected under 35 U.S.C. 112, first paragraph, as containing subject matter which was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention.

Claim 88 is drawn to a method of preparing a mutant yeast that produces the glycoprotein of formula (IV) by disrupting MNN1 gene, MNN4 gene and OCH1 gene in a wild-type yeast and transforming the resulting yeast with a polynucleotide encoding an α-mannosidase I and a polynucleotide encoding a GnT-I. Claims 92-93 limit claim 88 to specific auxotrophic mutations. Claim 94 is drawn to a method of preparing a mutant yeast that produces the glycoprotein of formula (IV) by disrupting MNN1 gene, MNN4

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gene and OCH1 gene in a wild-type yeast and transforming the resulting yeast with a polynucleotide encoding an Aspergillus saitoi α-mannosidase I and a polynucleotide encoding a GnT-I. These claims are drawn to a method of (A) mutagenizing any yeast to disrupt MNN1, MNN4 and OCH1 genes by any methods and (B) transforming said mutated yeast with a polynucleotide encoding any α-mannosidase I, including any recombinants, variants and mutants, and a polynucleotide encoding any GnT-I, including any recombinants, variants and mutants. Therefore, the claims are drawn to a method of mutagenizing any yeast that by disrupting a genus of MNN1, MNN4 and OCH1 genes and transforming said yeast with a polynucleotide encoding a genus of any α-mannosidase having any structure and a genus of GnT-I having any structure. There is insufficient descriptive support for using the above genus with respect to the yeast as well as α-mannosidase and GnT-I. The specification only describes a method of preparing a Saccharomyces cerevisiae mutant by disrupting MNN1, MNN4 and OCH1 genes normally present in S. cerevisiae with selection markers recited in claim 92 and transforming the resulting S. cerevisiae mutant with a polynucleotide encoding α-mannosidase I isolated from A. saitoi and a polynucleotide encoding a rat GnT-I (cloned by Yoshida et al – form PTO-1449). This one example is not enough and does not constitute a representative number of all the species to describe a method of mutagenizing any or all yeast using any method and transforming the resulting yeast with a polynucleotide encoding any or all α-mannosidase I and GnT-I, including any or all mutants, variants and recombinants. Further, there is no evidence on the record of the relationship between the structure of any or all yeast and the structure of a

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polynucleotide encoding an *A. saitoi* α-mannosidase I and the structure of any or all recombinants, variants and mutants of a polynucleotide encoding any α-mannosidase I or the structure of a polynucleotide encoding a rat GnT-I and the structure of a polynucleotide encoding any or all recombinants, variant or mutants of any GnT-I. Therefore, the specification fails to describe a representative species of mutagenizing a genus of yeast by disrupting a genus of MNN1, MNN4 and OCH1 genes by any methods and transforming the resulting yeast with a polynucleotide encoding a genus of α-mannosidase I and a genus of GnT-I.

Given this lack of description of the representative species encompassed by the genus of the claims, the specification fails to sufficiently describe the claimed invention in such full, clear, concise, and exact terms that a skilled artisan would recognize that applicants were in possession of the inventions of claims 88 and 92-94.

Applicant is referred to the revised guidelines concerning compliance with the written description requirement of U.S.C. 112, first paragraph, published in the Official Gazette and also available at www.uspto.gov http://www.uspto.gov.

In response to the previous Office Action, applicants have traversed the above rejection.

Applicants argue that the instant claims are fully described because the claimed method can be implemented with any yeast N-linked oligosaccharides contain a common mannose type sugar chain structure. Examiner respectfully disagrees. The rejection is not based on whether any or all yeast are able or unable to for the sugar chain of formula (IV), but that the specification does not describe mutagenizing any or

all yeast by disrupting MNN1, MNN4 and OCH1 genes. In order to practice the claimed method, one having ordinary skill in the art must be able to disrupt MNN1, MNN4 and OCH1 genes in any or all yeast, which entails knowing the structure of the MNN1, MNN4 and OCH1 genes. However, description of only Saccharomyces cerevisiae wherein MNN1, MNN4 and OCH1 genes normally present in said yeast are disrupted with selection markers recited in claim 92 is not enough to describe a genus comprising any or all yeast wherein MNN1, MNN4 and OCH1 genes normally present in said yeast or exogenous/recombinant forms of said genes, including any or all mutants or variants thereof, wherein said genes are disrupted by any methods. When there is substantial variation within the genus, one must describe a sufficient variety of species to reflect the variation within the genus, which applicants have failed to do. As discussed above, the written description requirement for a claimed genus may be satisfied through sufficient description of a representative number of species by disclosure of relevant, identifying characteristics, i.e., structure or other physical and/or chemical properties, by functional characteristics coupled with a known or disclosed correlation between function and structure.

Applicants also argue that the Lilly decision is not applicable to the present claims because in Lilly, the claims at issue were to genes while the present invention is drawn to a method of preparing a mutant yeast. Examiner respectfully disagrees. As discussed above, in order to practice the claimed method, one having ordinary skill in the art must be able to disrupt MNN1, MNN4 and OCH1 genes in any or all yeast, which entails knowing the structure of the MNN1, MNN4 and OCH1 genes. A description of

Saccharomyces cerevisiae wherein MNN1, MNN4 and OCH1 genes normally present in said yeast are disrupted with selection markers recited in claim 92 is not enough to describe a genus comprising any or all yeast wherein MNN1, MNN4 and OCH1 genes normally present in said yeast or recombinant forms of said genes, including any or all mutants or variants thereof, wherein said genes are disrupted by any methods. When there is substantial variation within the genus, one must describe a sufficient variety of species to reflect the variation within the genus, which applicants have failed to do. As discussed above, the written description requirement for a claimed genus may be satisfied through sufficient description of a representative number of species by disclosure of relevant, identifying characteristics, i.e., structure or other physical and/or chemical properties, by functional characteristics coupled with a known or disclosed correlation between function and structure.

Further, the claims are also drawn to a method of using a genus of any α-mannosidase having any structure and a genus of GnT-I having any structure, including any or all recombinants, variants or mutants. As discussed above, the written description requirement for a claimed genus may be satisfied through sufficient description of a representative number of species by disclosure of relevant, identifying characteristics, i.e., structure or other physical and/or chemical properties, by functional characteristics coupled with a known or disclosed correlation between function and structure. A description of transforming *Saccharomyces cerevisiae* with polynucleotide encoding α-mannosidase I isolated from *A. saitoi* and a polynucleotide encoding a rat GnT-I is not enough to describe a genus comprising any or all α-mannosidase and

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genus comprising any or all GnT-I, including any or all mutants or variants thereof.

When there is substantial variation within the genus, one must describe a sufficient variety of species to reflect the variation within the genus, which applicants have failed to do.

Hence the rejection is maintained.

Claims 88 and 92-94 remain rejected under 35 U.S.C. 112, first paragraph, because the specification, while being enabling for a method of preparing a mutant *Saccharomyces cerevisiae* by disrupting MNN1, MNN4 and OCH1 genes normally present in *S. cerevisiae* with selection markers recited in claim 92 and transforming the resulting mutant *S. cerevisiae* with a polynucleotide isolated from *Aspergillus saitoi* and encoding an α-mannosidase I and a polynucleotide isolated from rat and encoding a rat GnT-I, does not reasonably provide enablement for a method of preparing any mutant yeast that produces the glycoprotein of formula (IV) by disrupting any or all MNN1, MNN4 and OCH1 genes and transforming the resulting mutant yeast with a polynucleotide encoding any α-mannosidase I from any source, including recombinants, variants and mutants, and a polynucleotide encoding any GnT-I, including recombinants, variants and mutants. The specification does not enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the invention commensurate in scope with these claims.

Factors to be considered in determining whether undue experimentation is required are summarized in <u>In re Wands 858 F.2d 731, 8 USPQ2nd 1400 (Fed. Cir.</u>

1988). They include (1) the quantity of experimentation necessary, (2) the amount of direction or guidance presented, (3) the presence or absence of working examples, (4) the nature of the invention, (5) the state of the prior art, (6) the relative skill of those in the art, (7) the predictability or unpredictability of the art, and (8) the breadth of the claims.

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Claim 88 is drawn to a method of preparing a mutant yeast that produces the glycoprotein of formula (IV) by disrupting MNN1 gene, MNN4 gene and OCH1 gene in a wild-type yeast and transforming the resulting yeast with a polynucleotide encoding an α-mannosidase I and a polynucleotide encoding a GnT-I. Claims 92-93 limit claim 88 to specific auxotrophic mutations. Claim 94 is drawn to a method of preparing a mutant yeast that produces the glycoprotein of formula (IV) by disrupting MNN1 gene, MNN4 gene and OCH1 gene normally present in a wild-type yeast and transforming the resulting yeast with a polynucleotide encoding an Aspergillus saitoi α-mannosidase I and a polynucleotide encoding a GnT-I. These claims are drawn to a method of (A) mutagenizing any yeast, (B) disrupting MNN1, MNN4 and OCH1 genes by any methods and (C) transforming said mutant yeast with a polynucleotide encoding any αmannosidase I, including any recombinants, variants and mutants, and a polynucleotide encoding any GnT-I, including any recombinants, variants and mutants. Therefore, the claims are drawn to a method of mutagenizing any yeast by disrupting MNN1, MNN4 and OCH1 genes having any structure in any yeast using any methods and transforming said yeast with a polynucleotide encoding any α-mannosidase having any structure I and any GnT-I having any structure.

The scope of the claims is not commensurate with the enablement provided by the disclosure with regard to the extremely large number of yeasts wherein any MNN1, MNN4 and OCH1 genes are disrupted using any methods and wherein the resulting yeast is transformed with polynucleotides encoding any or all variants, mutants and recombinants of any α-mannosidase I and any GnT-I broadly encompassed in the method of the claims. Since applicants have not shown that the claimed method applies to all or any yeast and since the amino acid sequence of a protein determines its structural and functional properties, predictability of which changes can be tolerated in a protein's amino acid sequence and obtain the desired activity requires a knowledge of and guidance and predictability of which specific yeast is ideal for the claimed method and which specific α-mannosidase and GnT-I is ideal to transform said mutated yeast and which amino acids in the protein's sequence, if any, are tolerant of modification and which are conserved (i.e. expectedly intolerant to modification), and detailed knowledge of the ways in which the proteins' structure relates to its function.

However, in this case the disclosure is limited to a method of preparing a Saccharomyces cerevisiae mutant by disrupting MNN1, MNN4 and OCH1 genes normally present in *S. cerevisiae* with selection markers recited in claim 92 and transforming the resulting S. cerevisiae mutant with a specific polynucleotide isolated from Aspergillus saitoi encoding α-mannosidase I and a polynucleotide isolated from rat encoding GnT-I (cloned by Yoshida et al – form PTO-1449), but provides no guidance with regard to a method of comprising the use of any yeast for disrupting any or all MNN1, MNN4 and OCH1 genes using any methods and transforming the resulting

mutant yeast with a polynucleotide encoding any α-mannosidase I and a polynucleotide encoding any GnT-I. It would require undue experimentation of the skilled artisan to make and use the agents in the claimed method. In view of the great breadth of the claim, amount of experimentation required to mutagenize any yeast, identify and make the polynucleotides encoding any α-mannosidase I and GnT-I, amount of experimentation required to disrupt any MNN1, MNN4 and OCH1 genes using any methods, the lack of guidance, working examples, and/or unpredictability of the art in predicting function from a polypeptide primary structure, the claimed invention would require undue experimentation. As such, the specification fails to teach one of ordinary skill how to use the full scope of the polynucleotides and yeasts encompassed by the claims.

While enzyme isolation techniques, recombinant and mutagenesis techniques and other related techniques are known, and it is routine in the art to screen for multiple strains, multiple substitutions or multiple modifications in a polypeptide as encompassed by the instant claims, the specific yeast strains required for the method and the specific amino acid positions within a protein's sequence where amino acid modifications can be made with a reasonable expectation of success in obtaining the desired activity/utility for transforming said mutant yeast are limited in any protein and the result of such modifications is unpredictable. In addition, with respect to the polynucleotides encoding mannosidase and GnT-I used for transforming mutant yeast, one skilled in the art would expect any tolerance to modification for a given protein to diminish with each further and additional modification, e.g. multiple substitutions.

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The specification does not support the broad scope of the claims which encompass a method of preparing any yeast by disrupting any or all MNN1, MNN4 and OCH1 genes using any methods and transforming the resulting mutant yeast with a polynucleotide encoding any α-mannosidase I and a polynucleotide encoding any GnT-I because the specification does not establish: (A) said method will be successful in any or all yeasts; (B) a universal method disrupt any MNN1, MNN4 and OCH1 genes in any yeast; (C) a rational and predictable scheme for selecting agents, techniques or methods with an expectation of disrupting any MNN1, MNN4 and OCH1 genes in any yeast; (D) the general tolerance of α-mannosidase I and GnT-I to modification and extent of such tolerance; (E) a rational and predictable scheme for selecting any yeast for disrupting MNN1, MNN4 and OCH1 genes and transforming the resulting yeast with a polynucleotide encoding any α-mannosidase I and GnT-I with an expectation of making a mutant yeast that produces the glycoprotein of formula (IV); and (F) the specification provides insufficient guidance as to which of the essentially infinite possible choices is likely to be successful.

Thus, applicants have not provided sufficient guidance to enable one of ordinary skill in the art to make and use the claimed invention in a manner reasonably correlated with the scope of the claims broadly including a method of preparing any mutant yeast by disrupting any or all MNN1, MNN4 and OCH1 genes using any methods and transforming the resulting mutant yeast with a polynucleotide encoding any α-mannosidase I and a polynucleotide encoding any GnT. The scope of the claims must bear a reasonable correlation with the scope of enablement (*In re Fisher*, 166 USPQ 19

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24 (CCPA 1970)). Without sufficient guidance, determination of yeasts suitable for the above method and polynucleotides encoding an α-mannosidase I and GnT- having the desired biological characteristics is unpredictable and the experimentation left to those skilled in the art is unnecessarily, and improperly, extensive and undue. See *In re Wands* 858 F.2d 731, 8 USPQ2nd 1400 (Fed. Cir, 1988).

In response to the previous Office Action, applicants have traversed the above rejection.

Applicants argue that the instant claims are fully enabled because the claimed method can be implemented with any yeast N-linked oligosaccharides contain a common mannose type sugar chain structure. Examiner respectfully disagrees. The rejection is not based on whether any or all yeast are able or unable to for the sugar chain of formula (IV), but that the specification does not enable mutagenizing any or all yeast by disrupting MNN1, MNN4 and OCH1 genes, normally present in said yeast or exogenous/recombinant forms of said genes, including any or all mutants or variants thereof. In order to practice the claimed method, one having ordinary skill in the art must be able to disrupt MNN1, MNN4 and OCH1 genes in any or all yeast, which entails knowing the structure of the MNN1, MNN4 and OCH1 genes. However, the specification only teaches *Saccharomyces cerevisiae* wherein MNN1, MNN4 and OCH1 genes normally present in said yeast are disrupted with selection markers recited in claim. Therefore, it would require undue experimentation of the skilled artisan to make the claimed variants and mutants of any or all yeast wherein any or all MNN1, MNN4

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and OCH1 genes, normally present in said yeast or exogenous/recombinant forms of said genes, including any or all mutants or variants thereof, are disrupted.

Further, the claims are also drawn to a method of using any α-mannosidase having any structure and any GnT-I having any structure, including any or all recombinants, variants or mutants. It would require undue experimentation of the skilled artisan to make and use the claimed variants and mutants of any or all α mannosidase having and any or all GnT-I, including any or all recombinants, variants or mutants thereof. As discussed above, predictability of which changes can be tolerated in a protein's amino acid sequence and obtain the desired activity requires a knowledge of and guidance and predictability of which specific yeast is ideal for the claimed method and which specific a-mannosidase and GnT-I is ideal to transform said mutated yeast and which amino acids in the protein's sequence, if any, are tolerant of modification and which are conserved (i.e. expectedly intolerant to modification), and detailed knowledge of the ways in which the proteins' structure relates to its function. It is this specific guidance that applicants do not provide. Without specific guidance, those skilled in the art will be subjected to undue experimentation of making and testing each of the enormously large number of mutants that results from such experimentation.

Hence the rejection is maintained.

Claim Rejections - 35 USC § 102

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The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless -

(b) the invention was patented or described in a printed publication in this or a foreign country or in public use or on sale in this country, more than one year prior to the date of application for patent in the United

Claims 88 and 92-94 remain rejected under 35 U.S.C. 102(b) as being anticipated by Chiba et al.

Claims 88 and 92-94 are drawn to a method of preparing a mutant yeast that produces the glycoprotein of formula (IV) by disrupting a MNN1, MNN4 and OCH1 genes using the selection markers recited in claim 92 and introducing a polynucleotide encoding α-mannosidase I, such as that of A. saitoi, and a polynucleotide encoding GnT-I into the resulting mutant yeast.

Chiba et al. (form PTO-1449) discloses a method of preparing a mutant yeast that produces the glycoprotein of formula (IV) by transforming a S. cerevisiae comprising $\Delta mnn1\Delta mnn4\Delta och1$ triple mutant with a polynucleotide encoding an A. saitoi α-mannosidase I and a polynucleotide encoding GnT-I into (Figure 1 and pages 26299-26300). The S. cerevisiae Δ mnn1 Δ mnn4 Δ och1 mutant was prepared by disrupting the MNN1, MNN4 and OCH1 genes with selection markers as recited in claim 92. Therefore, the reference of Chiba et al. anticipates claims 88 and 92-94.

In response to the previous Office Action, applicants have traversed the above rejection.

Applicants argue that the present invention is not anticipated by Chiba et al. because introduction of the GnT-I gene into the mutant yeast wherein MNN1, MNN4

and OCH1 genes are disrupted have not be carried out in the laboratory. Examiner respectfully disagrees. Although Chiba et al. has not actually introduced a GnT-I gene into the mutant S. cerevisiae in laboratory settings, Chiba et al. discloses a "Strategy for genetic manipulation of S. cerevisiae" (description for Figure 1). The definition for "strategy" is "careful plan or method". In Figure 1, Chiba et al. clearly shows a method for making a "genetically manipulated S. cerevisiae" by disrupting Δ mnn1Δ mnn4 Δ och1 and introducing α-mannosidase I and GnT-I genes, which are normally found in mammals and not in yeast, into said yeast to produce the yeast-hybrid complex glycoprotein. MPEP 2131 states "A claim is anticipated only if each and every element as set forth in the claim is found, either expressly or inherently described, in a single prior art reference." In the instant application, Figure 1 of Chiba et al. discloses each and every element set forth in the claims, as discussed above. Further, the claims are not drawn to a mutant yeast transformed with GnT-I, but the claims are drawn to a method of making said mutant yeast, which is clearly taught by Chiba. Also, Figure 1 outlining the method of Chiba et al. is enabling because Chiba et al. teaches yeast wherein Δ mnn1 Δ mnn4 Δ och1 are disrupted and successful expression of both α mannosidase I and GnT-I in yeast (Pages 26300 and page 26303, last paragraph).

Applicants also argue that the claims are not anticipated by Chiba et al. since there is no description concerning whether or not the Golgi retention signal is necessary with respect to the GnT-I gene in the method of Chiba et al. and since at the time the instant invention was filed, it was common knowledge for a person skilled in the art to introduce the GnT-I gene without the targeting signal. Examiner respectfully disagrees.

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The claims do not recite any limitation of whether the GnT-I gene has a targeting signal or not. Although the claims are interpreted in light of the specification, limitations from the specification are not read into the claims. *In re Van Geuns*, 988 F.2d 1181, 26 USPQ2d 1057 (Fed. Cir. 1993).

Hence the rejection is maintained.

Conclusion

None of the claims are allowable.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Yong Pak whose telephone number is 571-272-0935. The examiner can normally be reached 6:30 A.M. to 5:00 P.M. Monday through Thursday.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ponnathapu Achutamurthy can be reached on 571-272-0928. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Any inquiry of a general nature or relating to the status of this application or proceeding should be directed to the receptionist whose telephone number is 571-272-1600.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll free).

Yong D. Pak

Patent Examiner 1652